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Nano sized ocular drug delivery system - an overview

R.Vijaya Muthu Manikandar*, Dharmaraj.M, Harivignesh.D, Subashini.B, Damemumdaric.A

Department of Pharmaceutics, Sri Ram Nallamani Yadava College of Pharmacy, Nallamani Nagar, Kodikurichi, Tenkasi-627804, Tamilnadu, India

ABSTRACT

Drug delivery to the eye has remained as one of the most challenging research for formulation scientists as ocular drug delivery system requires a series of specified characteristics according to the physiological structure of the eye. Eye is a unique and challenging organ for therapeutic drug delivery on to the surface as well as in the interior part of the ocular structure. The drug may bind to tear proteins and conjunctival mucin to treat the local ophthalmic diseases, liquid eye drop is the most desirable dosage form when considering convenience of administration and clinical compliance of the patients. These conventional dosage forms account for nearly 90% of the currently available marketed formulations owing to their simplicity and good acceptance by patients. However, conventional eye drops, most of which are present in the solution form, usually have quite a limited therapeutic efficacy due to the low bioavailability. The use of nano-approaches like nano-suspensions, nanoparticles, Nano emulsion, niosomes and liposomes has led to the solution of various solubility and permeability related problems of poorly soluble drugs like dexamethasone, cyclosporin, dorzolamide, gancyclovir and many more drugs. This review includes various nano sized formulations used to achieve prolonged contact time of drugs with the cornea and increase their bioavailability. Ophthalmic drug delivery is one of the most interesting and challenging endeavors facing the pharmaceutical scientist.

Keywords: Ocular drug delivery nano-suspensions, Nanoparticles, Nano emulsion, Niosomes and liposomes

INTRODUCTION

The development of drug delivery approach for the transportation of drug in a bio-available and safe manner to the target site is now becoming an exceedingly important area of bio-pharmaceutical researches. To be sure, a large number of novel drug delivery technologies surface every year and every segment of body part has been attempted as a potential target for the site of action. As a consequence, various smart drug delivery technologies with significant outcomes have been reported for BCS class-II and class-IV drugs,

Author for Correspondence:

R.Vijaya Muthu Manikandar*
Department of Pharmaceutics,
Sri Ram Nallamani Yadava College of Pharmacy,
Nallamani nagar, Kodikurichi,
Tenkasi-627804, Tamilnadu, India

peptides, proteins, etc. Further, among novel drug technologies, delivery advancement nanotechnology in formulation development for bio-degradable, non-biodegradable, nanoemulsions, nanoparticles, vesicular systems, implants, bioadhesive systems, etc are currently under intensive exploratory studies [1]. Typically, less than 5% of the drug applied penetrates the cornea/sclera and reaches the intraocular tissue, with the major fraction of the dose applied often absorbed systemically through the conjunctiva and nasolacrimal duct. On the other hand, corneal and conjunctival epithelia of human eye, along with the tear film, construct a compact barrier preventing the drug absorption into the intraocular area. This can result in low bioavailability and undesirable systemic side effects [2]. In this review, we presented the application of various nanoapproaches in the field of topical ophthalmic drug delivery attempted by numerous investigators over the last decade. Review also enlightens the amalgamation of mucoadhesive characteristics with nanotechnology for the enhancement of corneal residency and bioavailability. Furthermore, topical ocular delivery research is also summarized in the initial of the article.

CONVENTIONAL APPROACH

Conventional dosage forms such as solutions, suspensions and ointments account for almost 90% of the currently accessible ophthalmic formulations on the market [2]. They offer some advantages such as their ease of administration by the patient, ease of preparation and low production costs. However, there are also significant disadvantages, especially with the use of conventional solutions, including the very short contact time with the ocular surface and the fast nasolacrimal drainage, both leading to poor bioavailability of the drug. Nevertheless, conventional eye drops remain the most commonly used dosage forms in ocular delivery.

Solutions

The reasons for choosing solutions over other dosage forms include their favorable cost, simplicity of formulation development and production, and high acceptance by the patients [3]. However, they also exhibit major drawbacks, such as rapid and extensive pre-corneal loss, the high

absorption via the conjunctiva and nasolacrimal duct leading to systemic side effects, as well as increased instillation frequency resulting in low patient compliance. Some of these problems have been reduced by the addition of viscosity-enhancing agents such as cellulose derivates, which are believed to increase the viscosity of the preparation and consequently reduce the drainage rate.

Suspensions

Suspensions of the micronized drug [<5µm] in a suitable aqueous vehicle are formulated, where the active compound is water insoluble. It is assumed that the drug particles remain in the conjunctival sac, thus promoting a sustained release effect [4]. According Davies to topical ophthalmic suspensions have a number of limitations [5]. They need to be adequately shaken before use to ensure correct dosing, and the amount of drug required to achieve therapeutic benefit, only a moderate increase in bioavailability, rendering suspensions expensive in terms of their production costs [6].

Ointments

Ointments generally consist of a dissolved or dispersed drug in an appropriate vehicle base. They are the most commonly used semisolid preparations as they are well tolerated, fairly safe and increase the ocular bioavailability of the drug. On application, ointment breaks up into small oily droplets that remain in the cul-de-sac as a drug depot. The drug eventually gets to the ointment-tear interface due to the shearing action of the eyelids [7].

Viscosity enhancing systems

In order to reduce the lachrymal clearance of ophthalmic solutions, various polymers have been added to increase the viscosity of the conventional eye drops. Among the range of hydrophilic polymers investigated in the area of ocular drug delivery are polyvinyl alcohol [PVA] and polyvinyl pyrrolidone [PVP], cellulose derivatives such as methylcellulose and polyacrylic acids [Carbopols®]. [8]

In situ gelling systems

In situ gelling systems are viscous polymer-based liquids that exhibit sol-to-gel phase transition

on the ocular surface due to change in a specific physico-chemical parameter [ionic strength, temperature or pH]. They are highly advantageous over preformed gels as they can be easily instilled in liquid form, but are capable of prolonging the residence time of the formulation on the surface of the eye due to gelling [9].

NANOMEDICINE APPROACH

of nano-approaches like nano-The use suspensions, nanoparticles, nanoemulsion. niosomes and liposomes has led to the solution of various solubility and permeability problems of poorly soluble drugs dexamethasone, cyclosporin, dorzolamide, gancyclovir and many more [10]. Drugs can also be targeted to ocular tissue to allow region specific delivery and minimize side effects to other organs [11]. Besides this, depending on their particle charge, surface properties and relative hydrophobicity, nanoparticles can be designed for successfully overcoming corneal barriers. In addition to these points, encapsulation of drug in nanoparticles, nanospheres, liposomes etc, can also provide stability to the drug along with prolonged exposure of the drug by controlled release behaviour [10].

The use of nano-approaches like nanonanoparticles, suspensions, nanoemulsion, niosomes and liposomes has led to the solution of various solubility and permeability related problems of poorly soluble drugs like dexamethasone, cyclosporin, dorzolamide, gancyclovir and many more. Table-1 summarises the recent works on nano-approaches for drug delivery used in ophthalmic research.

Nanoparticles [NPS]/ Nanospheres/ Nanocapsules

Nanoparticles are sub-microscopic, colloidal system consisting of macromolecular substances that vary in size from 10 nm to 1000 nm. The drug may be dissolved, entrapped, adsorbed, attached or encapsulated into the nanoparticle matrix. Depending on the method of preparation, nanoparticles, nanospheres or nanocapsules can be obtained with different properties and release profile for the encapsulated drugs [10].

Chitosan Nanoparticles

Natural cationic polymer chitosan [CH] nanocarriers have attracted a great deal of attention because of its unique properties, such as acceptable biocompatibility and biodegradability. Chitosan [CH] is a cationic polysaccharide able to gel when it comes in contact with specific multivalent polyanions, such as sodium tripolyphosphate [TPP]. Nanoparticles are spontaneously formed upon mixing of CH and TPP solutions, through the formation of inter- and intramolecular linkages between the phosphate groups of TPP and the amino groups of CH [12].

Albumin Nanoparticles

Ganciclovir [GCV] loaded albumin nanoparticles for the treatment of cytomegalovirus retinitis. Its in-vitro studies indicated a burst release of the drug in 1 h, which continued in a sustained manner for 5 days and continued for almost 30 days. The result demonstrated its controlled delivery for longer period [13].

Gelatin Nanoparticles

Due to its biocompatible and biodegradable nature, its nanoparticles were studied as nano-sized ocular drug carrier. The developed formulations were studied for various characteristics like particle size, zeta potential, encapsulation efficiency, in vitro drug release. All particle sizes of pilocarpine and hydrocortisone loaded gelatin nanoparticles were obtained in the range 300-500 nm and 110-220 nm, respectively. The average zeta potential value for particles prepared at pH 6 equals -6.95 mV, for spheres prepared at pH 4 it amounts to -6.10 mV. Consequently, there is no significant effect of the preparation pH on zeta potential value of the particles obtained. But in case of hydrocortisone, zeta potential was found to be between -4 to -12 mV that showed significant influence of gelatin on the zeta potential value [14]

Alginate Nanoparticles

The potential of sodium alginate nanoparticles has been explored as novel vehicle for the prolonged topical ophthalmic delivery. Gatifloxacin-loaded nanoparticles studied and revealed a fast release during the first hour followed by a more gradual drug release during a 24h period by a non-Fickian diffusion process [15].

Polymethylmethacrylate [PMMA] nanoparticles

Polymethylmethacrylate [PMMA] nanoparticles are made up of in situ emulsion polymerisation technique. Briefly, monomeric methylmethacrylate is dissolved in water or phosphate—buffered saline or a solution or suspension of drugs or antigens in a concentration range of 0.1–1.5% [16]. Nanoparticles made of 66polyacrylamide or PMMA do not degrade either biologically or enzymatically, which makes them less attractive for topical nanodispersion, but act as good drug delivery carrier as for contact lenses and hydrogels.

Cellulose acetate phthalate Nanoparticles

Cellulose acetate phthalate has been used for in situ gelling of latex nanoparticles [17]. The preparation of these latex particles involves emulsification of polymer followed by solvent evaporation. This latex suspension, upon coming in contact with the lacrimal fluid at pH 7.2–7.4, gets converted to gel form in situ, thus increasing the residence time of instilled solution in the eye. But such system is associated with blurring of vision as a disadvantage.

PACA [polyacryl-cyanoacrylate] Nanoparticles

PACA [polyacryl-cyanoacrylate] particles properties of biodegradation possess bioadhesion that makes them interesting drug carriers for controlled ocular drug delivery and drug targeting. Wood et al showed that PACA nanoparticles were able to adhere to the corneal and conjunctival surfaces, due to the ability to entangle in the mucin matrix and form a noncovalent or ionic bond with the mucin layer of the conjunctiva, which represent their mucoadhesion property [11]. Polyalkylcyanoacrylate [PACA] nanoparticles and nanocapsules have been shown to improve and prolong the corneal penetration of hydrophilic and lipophilic drugs like Betaxolol and amikacin sulfate. Despite these attractive results, potential of the PACA nanoparticles is limited due to the disruption to the corneal epithelium cell membrane [18].

Poly caprolactone [PECL] nanoparticle/nanocapsules

Poly caprolactone [PECL] nanocapsules may serve as superior polymer systems for ocular drug delivery due to its biodegradable and biocompatible nature. Marchal-Heussler et al successfully demonstrated with infusion of compared nanoparticles prepared by using PACA, PECL nanoparticles [19]. It was shown that the PECL nanoparticles yielded the highest pharmacological effect. This was believed to be due to the agglomeration of these nanoparticles in the conjunctival sac. Nanocapsules of PECL were also tried for the topical ocular delivery of cyclosporin A [CYA]. It was found that poly-E-caprolactone coating increased the corneal levels of the drug by 5 times compared to the oily solution of the drug when administered to the cul-de-sac of fully awake New Zealand white rabbits [12].

Eudragitt® nanoparticle

Eudragit® polymer nanoparticle suspensions have been investigated as a carrier system for the ophthalmic release of nonsteroidal antiinflammatory drugs, such as ibuprofen and flurbiprofen [20]. These inert resins based particles are proposed as delivery systems to prolong the release and improve ocular availability of the drug. They are reported to be devoid of any irritant effect on cornea, iris, and conjunctiva and thus appear to be a suitable inert carrier for ophthalmic drug delivery.

Polybutylcyanoacrylate nanoparticle

Polybutylcyanoacrylate [PBCA] nanoparticle delivery system for pilocarpine nitrate has been evaluated in comparison to the solution of the drug for pharmacokinetic and pharmacodynamic aspects [18,21,22], successfully demonstrated the effect of loaded polybutylcyanoacrylate pilocarpine nanoparticles on evaluated aqueous humor drug levels and the intraocular pressure-lowering effects using three models [the water-loading model, the alpha-chymotrypsin model, and the betamethasone model] in rabbits. The miotic response was enhanced by about 33% while the miotic time increased from 180 to 240 minutes nanoparticles compared to the control solution. PEG-coated Acyclovir-loaded polyethyl-2cyanoacrylate [PECA] nanospheres prepared by emulsion polymerization technique showed increased drug levels in the aqueous humor compared to the free drug suspension in the rabbits [23]

PLGA Nanoparticles

Polylactide and polylactide-co-glycolide biopolymers in the molecular weight range of 3000–109,000 have been employed in the preparation of nanoparticulate systems for intravitreal administration of acyclovir [24]. PLGA is a biodegradable and biocompatible polymer that is hydrolytically degraded into non-toxic oligomer and monomer, lactic acid and glycolic acid [25].

Non-polymeric nanoparticles

Chen et al. developed a new system for local delivery of methazolamide to eye based on calcium phosphate [CaP] nanoparticles. The methazolamide loaded CaP nanoparticles were prepared through the formation of an iorganic core of CaP and further adsorption of the methazolamide. In vitro release studies demonstrated diffusion-controlled release of methazolamide from the CaP nanoparticles over a period of 4 hr. In in vivo studies, it was indicated that the intraocular pressure [IOP] lowering effect of the inorganic nanoparticles eye drops lasted for 18 h, which was significantly better than the effect of 1% brinzolamide eye drops [6 h] [26,27],

Microemulsion/Nanosuspension [MS/NS]

Nanoemulsions are defined as the dispersions of water and oil in the presence of combination of surfactant and co-surfactant [Smix] in a manner to reduce interfacial tension. On the basis of nature of dispersion and disperse phase, NEs were classified as: o/w, w/o & bicontinuous type. These systems are usually characterized by clear appearance, higher thermodynamic stability, small droplet size [< 200 nm], high drug solubility, drug reservoir for lipophilic and hydrophilic drugs.

Liposomes

Liposomes are the microscopic vesicles composed of one or more concentric lipid bilayers, separated by water or aqueous buffer compartments with a diameter ranging from 100 nm to 10 µm [28, 29]. Liposomes offer advantages over most ophthalmic delivery systems in being completely biodegradable and relatively non-toxic. Another potential advantage of liposomes is their ability to come in an intimate contact with the corneal and conjunctival surfaces, thereby, increasing the probability of ocular drug absorption.

Niosomes

Like liposomes, in recent years, niosomes have been successfully studied for ophthalmic drug delivery as vesicular systems to provide controlled drug delivery, prolonged drug precorneal residence time and enhanced ocular bioavailability and prevention of metabolism of the drug by enzymes present at the tear/corneal surface [30]

Dendrimers

As per the definition given by **Sahoo** *et al.*, "Dendrimers are macromolecular compounds made up of a series of branches around a central core". Their nanosize, ease of preparation, functionalization and possibility to attach multiple surface groups render them suitable alternative vehicle for ophthalmic drug delivery" [31, 32, 33]

Solid lipid nanoparticles [SLN]

Solid lipid nanoparticles [SLN] are characteristically spherical particles with average diameter between 50 to 100nm [34]. SLNs are particularly advantageous in ocular drug delivery as they have the ability to enhance the ocular bioavailability of both hydrophilic and lipophilic drugs. Furthermore, they can be easily autoclaved for sterilization which is an important aspect of ocular administration for drug formulation [35].

Table 1: List of different nano-carriers explored by different investigator as possible topical ocular drug delivery carrier

Polymers / Bases/Surfactants based nano-carriers	Drug Encapsulated	Achievement	References
Niosomes – Modification of corneal peri	neability		_
Tween 60/Brij 35/Tween 80	Gentamicin	Controlled release, Enhanced	36

		corneal depositio	n.	
Span 60/ Cholesterol/ Chitosan	Timolol maleate	Controlled releas	e, Enhanced	37
Span 40/ Span 60/ Cholesterol	Acetazolamide	Enhanced permed	ation .	38
Sorbitan esters and poly-oxyethylene ulkyl ethers	Naltrexone Hydrochloride	Thermoresponsiv	U	39
Nanosuspensions – Endocytosis and mod	ification of cornea	ıl permeability		
Eudragit RS 100	Ibuprofen	Controlled releas	e	20
Eudragit RS 100R/ RL 100R	Flurbiprofen	Controlled releas	e	20.
Eudragit RS 100R/ RL 100	Cloricromene	Improved bioavai	ilability	40
PLGA/Poly[Lac[Glc-Leu]]	Diclofenac	Extended release	4	41
Pluronic F127	Timolol	High drug loadin	g	<i>4</i> 2
Sorbitan mono laurate oleate	Pilocarpine	Increased ocular	retention	43
Polyoxyethylenesorbitan monooleate				
Liposomes- modification of corneal per	meability			
Phosphatidyl choline / cholesterol		Puerarin	Increased ocular retention	
Phosphatidyl choline / cholesterol		Gatifloxacin	Improved bioavailabi	lity 4
Chitosan/\(\beta\)-glycerophosphate		Ofloxacin	Improved bioavailabi	lity 4
Phosphatidylcholine/ cholesterol		Acetazolamide	Increased ocular retention	2
Phosphatidylcholine/ cholesterol		Ganciclovir	Enhanced permeation	! 4
Dipalmitoylphosphatidyl choline /		Acyclovir	Enhanced permeation	! 4
Cholesterol/Dimethyl-dioctadecyl glycer	rol bromide		_	
Phosphatidylcholine/ cholesterol		Ciprofloxacin	Improved bioavailabi	lity 4
Submicron-sized liposomes		Edaravone	Inhibition of in vitro light-induced reactive oxygen species [ROS] production	
Chitosan coated liposomes		Cyclosporin A	Prolonged drug retention, enhanced drug permeation, and biocompatibility.	
Chitosan coated liposomes		Ciprofloxacin HCl	Improved ocular permeation	Š
N-trimethyl chitosan [TMC]-coated lipo	somes	Coenzyme Q[10]	Enhanced permeation	:
Phospholipids and cholesterol		Tacrolimus	Highly effective in suppressing the proce of autoimmune uveoretinitis	ess
Cationic and anionic surfactants		Calcein dye	Increases the permeability	2
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Ciprofloxacin

hydrochloride

Improved the ocular

bioavailability

36

 $L\hbox{-}\alpha\hbox{-}phosphatidylcholine, Cholesterol, Stearylamine and$

Dendrimers – Diffuses through paracellular pathway

dicetyl phosphate

Poly[amidoamine] [PAMAM]	Pilocarpine nitrate	Increased ocular	56
		retention	
Poly	Tropicamide	Increased ocular retention	56
[amidoamine] [PAMAM] anionic PANAM	D[+]-glucosamine	Synergistic	57
unone Phylin	and $D[+]$ -	immunomodulatory and	37
	glucosamine 6-	antiangiogenic	
	sulfate		
PANAM	anti-	better mechanical and	58
D. L. COLCA DIALNE E. L. C.	VEGF ODN	adhesion ability	
Polyester [PLGA, PLA] NPs –Endocytosis			
PLGA	Sparfloxacin	Increased ocular	25
DICA	Madad tonas	retention	50
PLGA PLGA	Methyl trypsin Flurbiprofen	Controlled release Enhanced penetration	59 60
PLGA	Dexametha zone	Sustained release	61
PLGA	Chloramphenicol	Prolonged release	62
PLA	Betamethasone	Targeting specific	63
		lesion; controlled	
		release	
PBCA NPs - Transcellular pathway or endocytosis			
PBCA	Cyclo-phosphamide	Prolonged release	64
PBCA	Pilocarpine	Prolonged release	65
Poly-ε-caprolactone NPs - Transcellular pathway or e	ndocytosis		
PECL	Indomethacin	Increased penetration	12
PECL	Carteolol	Pronounced decrease in IOP	66
Eudragit NPs – Endocytosis			
Eudragit RL 100	Amphotericin B	Controlled release	67
Eudragit RS 100	Piroxicam	Improved ocular	68
Ţ		delivery	
Chitosan NPs – Paracellular pathway			
Chitosan	BSA/Fluorescein	Good toleration	69
Chitosan	Indomethacin	Controlled release	70
Chitosan	Cyclosporine A	Increased	71
Albumin NDs Dhacomtosis		bioavailability	
Albumin NPs – Phagocytosis			
Albumin	Ganciclovir	Prolonged residence in eye	72
Albumin	Pilocarpine	Prolonged residence in	21
	•	eye	
Albumin	Aspirin	Prolonged residence in	73
		eye	
Albumin	Hydrocortisone	Targeting precorneal	21
		area	

Gelatin NPs			
Gelatin	Pilocarpine	High encapsulation	57
Gelatin	Hydrocortisone	Sustained release	57
PEG NPs - Phagocytosis			
PEG	Tamoxifen	Localization of NPs	74
Modified NPs			
Eudragit RS 100/ RL 100/ PLGA	Ciprofloxacin	Prolonged drug release	75
Eudragit RS 100/ RL 100/ hyaluronic acid	Gatifloxacin, prednisolone	Prolonged drug release	76
Chitosan/ alginate	Carboplatin	Sustained release	77
Chitosan/ hyaluronic acid	Dorzolamide	Reduction in IOP level	<i>78</i>
Chitosan/ sodium alginate	Gatifloxacin	Prolonged drug delivery	15
PLA/ PEG	Acyclovir	Effective	<i>79</i>
Deacetylated water-soluble chitosan	Flurbiprofen	Transcorneal penetration and precorneal retention	80
Chitosan/ poly [D, L-lactic acid] [PLA]	5-fluorouracil	Non-irritant, Diffusion controlled release, Improved bioavailability	81
Chitosan Oligosaccharides [COS]-coated NLC [nanostructured lipid carrier]	Flurbiprofen	Enhanced transcorneal penetration	82
poly- epsilon -caprolactone [PECL], chitosan [CS]-coated PECL, poly[ethyleneglycol][PEG]-coated PECL nanocapsules.	Fluorescent dye [rhodamine]	Improved biodistribution in eye	83
Solid Lipid Nanoparticle			
Stearicacid, Compritol [SLN-B] and poloxamer-188	Gatifloxacin	Physiologically tolerable	84
GMS/ Stearic acid	Methazolamide	Prolonged effect, better patient compliance	85
GMS/ poloxamer	Baicalin	Enhanced ocular bioavailability	86
Phospholipid	Timolol hydrogen maleate	Sustained permeation	87
Dynasan 116	Cyclosporine A	Increased residence time	88
Hexadecyl phosphate	Tobramycin	Improved Bioavailability	89
Glyceryl monostearate/ Poloxamer 188	Chloramphenicol	Improved drug entrapment efficiency and controlled drug release	90
Homolipid/ Phospholipid	Diclofenac sodium	Improved permeation	91

CURRENT CONCEPT

From last 20 years, efforts have been directed in the rational design of ocular drug delivery consisting of mucoadhesive nano-carriers. Thereafter, the plan was directed to generate nanocarriers with a hydrophilic coating with the idea of improving their stability and their interaction with the mucosa [92, 93, 83, 94].

It was proficient via optimization of nanocarriers ocular drug delivery to obtain long-lasting bioadhesion/residence time by the so-called 'mucoadhesive property' based on entrapment of particles in the ocular mucus layer and interaction of bioadhesive polymer chains with mucins [95]. Maintenance of the designed nanocarriers in the ocular delivery following topical application is thus decisive to accomplish unremitting drug release and prolonged therapeutic activity. Therefore, manufacture of nanocarriers from mucoadhesive materials is crucial for effective retention in ocular cul-de-sac [95].

Ocular mucoadhesion, exclusively, refers to the capability of certain polymers to hold on to the mucus layer casing the conjunctival and corneal surfaces of the eye by non-covalent bonds. Washout time of mucoadhesive polymeric system is reduced, since this depends on mucous turnover rate rather than lachrymal discharge turnover rate. Mucoadhesive polymer with plentiful hydrophilic functional group viz sulphate, hydroxyl, carboxyl, amide have found fundamental role in ocular drug delivery system owing to their adhesion property with precorneal/conjunctival mucin layer via noncovalent bonds, and remaining in place for as long as the mucin is available there. Using this concept, various investigators planned the cationic polymer Chitosan as a polymer of choice because of its unique properties, including good enough biodegradability and biocompatibility [96, 97, 15, 93, 98]. Furthermore, it was found that, CH increases cell permeability by affecting both paracellular and intracellular pathways of epithelial cells in a reversible manner without affecting cell viability or causing membrane wounds [96, 97, 15, 93, 981.

During the selection of bioadhesive polymer intended for ophthalmic drug delivery, the viscosity and wetting properties of polymer are considered. Viscosity measures the resistance to flow which

depends upon its molecular mass, concentration, temperature and shear stress. In Newtonian system, above a certain range of viscosity there is no real improvement of bioavailability and no further increase of residence contact time and blinking becomes a panic [99,100,101].

On the other hand, polymer showing non-Newtonian behaviour, when incorporated in formulation possessing pseudoplastic behaviour in which viscosity decreases with increasing shear rate [due to blinking and ocular movement], this results in significantly less resistance to blinking and demonstrates greater acceptance as compared to formulation possessing polymer exhibiting Newtonian flow [101,102]. The mucoadhesive properties of polyacrylic acid hydrogels and their ability to penetrate the mucin at the surface of the have been investigated eye extensively [103,104,105,106]. In the mean time, several other synthetic polymers have been examined for the fabrication of mucoadhesive nanocarriers for ocular delivery, for example Pignatello et al reported the formulation and evaluation of nanocarriers composed of Eudragit-RL100 with good ocular tolerance, and no inflammation or discomfort in the rabbit eye [20]. According to [75].positively charged nanoparticles could also be prepared when Eudragit RL100 was combined with PLGA.

Barbault-Foucher S et.al reported the Hyaluronic acid [HA] as a natural, nonirritating polysaccharide showing pseudoplastic behaviour with desirable ocular mucoadhesive properties. This group designed the novel ocular drug delivery system based on biodegradable nanospheres coated with a mucooadhesive polymer. The system was composed of a core of poly-e-caprolactone coated by corona of the bioadhesive HA molecule. In this investigation, this group proposed the non-covalent attachment of unmodified HA to the exterior of the nanoparticles. They use three approaches viz, i] coating the poly-e-caprolactone core during particle formation by chain entanglement with HA; ii] of preformed poly-e-caprolactone coating nanosystems by HA adsorption; and iii] coating of poly-e-caprolactone nanosystem by electrostatic interactions between negatively charged HA and a cationic surfactant used in the formulation [i.e., a cationic lipid, stearylamine, and a preservative usually used in ophthalmic formulation and absorption enhancer, benzalkonium chloride. The

results made known that HA was robustly attached to nanospheres that had been conferred with a positive charge by cationic surfactant, resulting in intact HA-coated nanospheres. However, like chitosan it does not possess permeability-enhancing properties [108]; in addition, the toxicity of stearylamine must be taken into consideration [95]. Investigator reported the use of gelatin nanoparticle based on its biocompatibility and biodegradability, since it was derived from the collagen obtained from stroma of the eye, and has been used in the ocular extensively in drug delivery [109,110,111]. Tailoring of nano-systems with positive surface functionalization was extensively explored in the recent research trend of ocular delivery technology. [112] reported the synthesis

and evaluation of positively charged phospholipids and cholesterols as components for liposomes. This group reported that some liposome preparations containing these synthetic lipid materials were found to be non-cytotoxic. Further, they observe that insertion of the positively charged lipid derivatives into the liposomes appreciably improved the ocular withholding compared with neutral or negatively charged liposomes in an unanaesthetized rabbit eye model, due to molecular association with polyanionic corneal and conjunctival surface mucoglycoproteins. The positively charged functionalized nano-ophthalmic carriers more particularly chitosan imparted positive charged nanosystems investigated in recent time is summarized in Table 2.

Table 2: List of mucoadhesive nano-systems investigated in recent years for enhanced ocular retention and bioavailability of drugs

Mucoadhesive	Therapeutic	Transport Mechanism	Efficacy	Reference
Nano-system	Drug/Biomolecule			
Chitosan nanoemulsions [nanocapsules]	Indomethacin	Endocytic, Transcellular.	Significant increase in drug availability in cornea and aqueous humour.	70
Chitosan coated PECL nanocapsules	Indomethacin	Interaction with corneal surface due to positive charge, transcellular pathway.	Significant increase in drug availability in cornea and aqueous humour.	113
Chitosan nanoparticles	Cyclosporin A	Improved interaction with the cornea and conjunctiva, transcellular pathway, specific affinity for some conjunctival cell.	significant increase in the CYA concentration in the cornea.	83
	proteins	Active transport, temperature dependent endocytosis.	Increased concentration in ocular tissue.	69
			expression of the encoded protein upon their incubation with corneal and conjunctival epithelial cells.	
	pDNA			114
Chitosan coated	PDIM	Effective interaction with	Significant increase in	115
PLA nanoparticles	Rapamycin	ocular mucosa	allograft survival.	110

Self-assembled Chitosan nanoparticles	Prednisolone	Transcellular pathway.	Uniform distribution and improved retention on ocular surface, increase in concentration in aqueous humour.	116
Chitosan polysaccharide nanoparticles	pDNA	Interact with the HA-receptor CD44, expressed in ocular cell lines.	significant protein expression corneal and conjunctival epithelial cells.	114
Chitosan lipid nanoparticles	Flurbiprofen	Transcorneal penetration, endocytosis.	Improved Precorneal retention.	80
Chitosan Nanostructured lipid carrier	Flurbiprofen	Transcorneal penetration.	Delayed clearance from ocular surface.	82
chitosan-coated liposomes	Cyclosporin A	Transcellular pathway, positive charge interaction with mucin.	concentrations increased in cornea, conjunctiva, and sclera.	51
Chitosan Liposomes	Ciprofloxacin hydrochloride	Diffusion.	enhanced antimicrobial activity against both Grampositive and Gram-negative bacteria.	52
N-trimethyl chitosan [TMC]-coated liposomes	Coenzyme Q[10]	Transepithelial transport.	elevated the cell viability and reduced the oxidative damage.	53
Chitosan coated niosomes	Timolol	Paracellular pathway, corneal retention.	Sustained control of the intraocular pressure.	117

Overall, the promising results illustrated in the literatures point towards the acceptance of nanotechnological carry as future nanomedicine for topical ocular administration bioactives. In addition, surface functionalized with positive charge nanomedicine has emerged as an extremely promising candidate, given their improved interaction with the ocular surface and hence, enhanced ocular residence with prolonged delivery of the carried bioactives in ophthalmology.

CONCLUSION

Development of nano- sized novel formulations is worthwhile in such cases of ophthalmic delivery as they are expected to prolong the pre-ocular

increase the ocular retention and bioavailability.Drug enclosed in the vesicles and oily nano-droplets allows for an improved partitioning and transport through the cornea. Moreover, vesicles offer a promising avenue to fulfil the need for an ophthalmic drug delivery system that has the convenience of a drop, but will localize and maintain drug activity at its site of action [118, 31]. Finalythis review conclude that nano sized formulation, [nanoemulsion, niosome, etc.,] were achieved higher inner ocular tension and minimize systemic drug absorption. Hence we proposed novel nano sized formulation were suitable for prolonged delivery of drug in ophthalmology.

REFERENCES

- [1]. Akhter S, Talegaonkar S, Khan ZI, Jain GK, Khar RK, Ahmad FJ. Assessment of ocular pharmacokinetics and safety of Ganciclovir loaded Nanoformulation. Biomed Nanotechnol. 7, 2011, 144-145.
- [2]. Lang JC. Ocular drug delivery conventional ocular formulations. Adv. Drug Deliv. Rev. 16, 1995, 39-43
- [3]. Fitzgerald P, Hadgraft J, Kreuter J, Wilson CG. A γ-scintigraphic evaluation of microparticulate ophthalmic delivery systems: liposomes and nanoparticles. Int J Pharm. 40, 1987, 81-84.
- [4]. Kupferman A, Pratt MV, Suckewer K, Leibowitz HM. Topically applied steroids in corneal disease, the role of drug derivative in stromal absorption of dexamethasone. Arch. Ophthalmol. 91, 1974, 373-376.
- [5]. Davies NM. Biopharmaceutical Considerations in Topical Ocular Drug Delivery. Clin Exp Pharmacol Physiol. 27, 2000, 558-562.
- [6]. Davies NM, Wang G, Tucker IG. Evaluation of a hydrocortisone/hydroxypropyl-β-cyclodextrin solution for ocular drug delivery. Int. J. Pharm. 156, 1997, 201-209.
- [7]. MacKeen DL. Aqueous formulations and ointments. Int Ophthalmol Clin. 20, 1980, 79-92.
- [8]. Chrai SS, Robinson JR. Ocular evaluation of methylcellulose vehicle in albino rabbits. J Pharm Sci. 63, 1974, 1218-1223.
- [9]. Krauland AH, Leitner VM, Bernkop-Schnürch A. Improvement in the in situ gelling properties of deacetylated gellan gum by the immobilization of thiol groups. J Pharm Sci. 92, 2003, 1234-1241.
- [10]. Sahoo SK, Labhasetwar V. Nanotech approaches to drug delivery and imaging. Drug Discov Today 8, 2003, 1112–1120.
- [11]. Wood RW, Li VHK, Kreuter J, Robinson JR. Ocular disposition of poly-hexyl-2-cyano [3-14C] acrylate nanoparticles in the albino rabbit. Int J Pharm 23, 1985, 175-183.
- [12]. Calvo P, Sanchez A, Martinez J, Lopez MI, Calonge M, Pastor JC, Alonso MJ. Polyester Nanocapsules as new topical ocular delivery systems for cyclosporin A. Pharm Res. 13, 1996, 311-315.
- [13]. Merodio M, Arnedo A, Renedo MJ, Irache JM. Ganciclovir-loaded albumin nanoparticles: characterization and in vitro release properties. Eur J Pharm Sci 12, 2001, 251–259.
- [14]. Vandervoort J, Ludwig A. Ocular drug delivery: nanomedicine applications Nanomedicine [Lond]. 2, 2007, 11-21.
- [15]. Motwani SK, Chopra S, Talegaonkar S, Kohli K, Ahmad FJ, Khar RK. Chitosan-sodium alginate nanoparticles as submicroscopic reservoirs for ocular delivery: formulation, optimisation and in vitro characterization, Eur. J. Pharm. Biopharm 68, 2008, 513–525.
- [16]. Kreuter J. Nanoparticles-Preparation and applications. In: Donbrow M. editors. Microcapsules and Nanocapsules in Medicine and Pharmacy. CRC Press Inc, Boca Raton; 1992, 126–143.
- [17]. Gurny R. Latex systems. In: Breimer DD, Speiser P. editors. Topics in Pharmaceutical Sciences. Elsevier Science Publishers, Amsterdam, 1983, 277–288.
- [18]. Zimmer A, Kreuter J, Robinson JR.. Studies on the transport pathway of PBCA nanoparticles in ocular tissues. J Microencaps. 8, 1991, 497–504.
- [19]. Marchal-Haussler L, Fessi H, Devissaguet JP, Hoffman M, Maincent P. Colloidal drug delivery systems for the eye. A comparison of the efficacy of three different polymers: polyisobutylcyanoacrylate, polylactic-coglycolic acid, poly-epsilon-caprolactone. Pharm. Sci. 2, 1992, 98.
- [20]. Pignatello R, Bucolo C, Ferrara P, Maltese A, Puleo A, Puglisi G. Eudragit RS100® nanosuspensions for the ophthalmic controlled delivery of ibuprofen. Eur J Pharm Sci 16, 2002, 53-61.
- [21]. Zimmer A, Mutschler E, Lambrecht G, Mayer D, Kreuter J. Pharmacokinetic and Pharmacodynamic aspects of an ophthalmic pilocarpine nanoparticle-delivery-system. Pharm Res. 11, 1994, 1435-1442.
- [22]. Diepold R, Kreuter J, Himber J, Gurny R, Lee VH, Robinson JR, Saettone MF, Schnaudigel OE. Comparison of different models for the testing of pilocarpine eyedrops using conventional eyedropsas a novel depot formulation [nanoparticles]. Graefe's Arch. Clin Exp Ophthalmol 227, 1989, 188-193.
- [23]. Gurny R. Ocular therapy with nanoparticles. In: Guiot P, Couvreur P. editors. Polymeric Nanoparticles and Microspheres. CRC Press, Boca Raton; 1986, 127–136.

- [24]. Conti B, Bucolo C, Giannavola C, Puglisi G, Giunchedi P, Conte U. Biodegradable microspheres for the intravitreal administration of acyclovir: in vitro in vivo evaluation. Eur J Pharm Sci. 5, 1997, 287-293.
- [25]. Gupta H, Aqil M, Khar RK, Ali A, Bhatnagar A, Mittal G. Sparfloxacin-loaded PLGA nanoparticles for sustained ocular drug delivery. Nanomedicine. 6, 2010, 324-333.
- [26]. Chen R, Qian Y, Li R, Zhang Q, Liu D, Wang M, Xu Q. Methazolamide Calcium Phosphate Nanoparticles in an Ocular Drug Delivery System. Yakugaku Zasshi. 130, 2010, 419-424.
- [27]. Sheikpranbabu S, Kalishwaralal K, Venkataraman D, Eom SH, Park J, Gurunathan S. Silver nanoparticles inhibit vegf-and il-1 -induced vascular Permeability via src dependent pathway in porcine retinal Endothelial cells J Nanobiotechnology. 0, 2009, 7:8.
- [28]. Wadhwa S, Paliwal R, Paliwal SR, Vyas SP. Nanocarriers in ocular drug delivery: an update review. Curr Pharm Des. 15, 2009, 2724-2750.
- [29]. Davis JL, Gilger BC, Robinson MR. Novel approaches to ocular drug delivery. Curr Opin Mol Ther. 6, 2004, 195-205.
- [30]. Kaur IP, Garg A, Singla AK, and Aggarwal D. Vesicular systems in ocular drug delivery: an overview. Int J Pharm. 269, 2004, 1–14.
- [31]. Sahoo SK, Dilnawaz F, Krishnakumar S. Nanotechnology in ocular drug delivery. Drug Discov Today. 13, 2008, 144–151.
- [32]. Quintana A, Raczka E, Piehler L, Lee I, Myc A, Majoros I, Patri AK, Thomas T, Mulé J, Baker JR Jr. Design and function of a dendrimer-based therapeutic nanodevice targeted to tumor cells through the folate receptor. Pharm Res. 19, 2002, 1310–1316.
- [33]. Ihre HR, Padilla OL, Jesus D, Szoka Jr. FC, Frechet JM. Polyester dendritic systems for drug delivery applications: design, synthesis, and characterization. Bioconjug Chem 13, 2002, 443–452.
- [34]. Muller RH, Keck CM. Challenges and solutions for the delivery of biotech drugs--a review of drug nanocrystal technology and lipid nanoparticles. J Biotechnol. 113, 2004, 151-170.
- [35]. Seyfoddin A, Shaw J, Al-Kassas R. Solid lipid nanoparticles for ocular drug delivery. Drug Deliv. 17, 2010, 467-489.
- [36]. Abdelbary G. Ocular ciprofloxacin hydrochloride mucoadhesive chitosan-coated liposomes. Pharm Dev Technol. 16, 2011, 44-56.
- [37]. Kaur IP, Aggarwal D, Singh H, Kakkar S. Improved ocular absorption kinetics of timolol maleate loaded into a bioadhesive niosomal delivery system. Graefes Arch Clin Exp Ophthalmol. 248, 2010, 1467-1472.
- [38]. Guinedi AS, Mortada ND, Mansour S, Hathout RM. Preparation and evaluation of reverse-phase evaporation and multilamellar niosomes as ophthalmic carriers of acetazolamide. Int J Pharm. 306, 2005, 71–82.
- [39]. Abdelkader H, Ismail S, Kamal A, Alany RG. Preparation of niosomes as an ocular delivery system for naltrexone hydrochloride: physicochemical characterization. Pharmazie. 65(11), 2010, 811-817.
- [40]. Pignatello R, Ricupero N, Bucolo C, Maugeri F, Maltese A, Puglisi G. Preparation and characterization of eudragit retard nanosuspensions for the ocular delivery of cloricromene. AAPS PharmSciTech. 7, 2006, E27.
- [41]. Agnihotri SM, Vavia PR Diclofenac-loaded biopolymeric nanosuspensions for ophthalmic application. Nanomedicine. 5, 2009, 90-95.
- [42]. Li CC, Abrahamson M, Kapoor Y, Chauhan A. Timolol transport from microemulsions trapped in HEMA gels. J Colloid Interface Sci 315, 2007, 297–306.
- [43]. Chan J, Maghraby GM, Craig JP, Alany RG. Phase transition water-in-oil microemulsions as ocular drug delivery systems: in vitro and in vivo evaluation. Int J Pharm. 328, 2007, 65-71.
- [44]. Deng Y, Xu J, Li X. Preparation and lacrimal pharmacokinetics of eye drops of puerarin liposomes in rabbit tears. Zhongguo Zhong Yao Za Zhi. 35, 2010, 301-304.
- [45]. Hosny KM. Ciprofloxacin as Ocular Liposomal Hydrogel. AAPS PharmSciTech. 11, 2010, 241-246.
- [46]. Hosny KM. Preparation and evaluation of thermosensitive liposomal hydrogel for enhanced transcorneal permeation of ofloxacin. AAPS PharmSciTech. 10, 2009, 1336-1342.
- [47]. Hathout RM, Mansour S, Mortada ND, Guinedi AS. Liposomes as an ocular delivery system for acetazolamide: in vitro and in vivo studies. AAPS Pharm SciTech. 8, 2007, 1.
- [48]. Shen Y, Tu J. Preparation and ocular pharmacokinetics of ganciclovir liposomes. Aaps J 9, 2007, E371–377.

- [49]. Chetoni P, Rossi S, Burgalassi S, Monti D, Mariotti S, Saettone MF. Comparison of Liposome-Encapsulated Acyclovir with Acyclovir Ointment: Ocular Pharmacokinetics in Rabbits. J Ocul Pharmacol Ther. 20, 2004, 169-177.
- [50]. Shimazaki H, Hironaka K, Fujisawa T, Tsuruma K, Tozuka Y, Shimazawa M, Takeuchi H, Hara H. Edaravone-loaded liposome eyedrops protect against light-induced retinal damage in mice. Invest Ophthalmol Vis Sci. 52, 2011, 7289-7297.
- [51]. Li N, Zhuang CY, Wang M, Sui CG, Pan WS. Low molecular weight chitosan-coated liposomes for ocular drug delivery: In vitro and in vivo studies. Drug Deliv. 19, 2012, 28-35.
- [52]. Mehanna MM, Elmaradny HA, Samaha MW. Ciprofloxacin Liposomes as Vesicular Reservoirs for Ocular Delivery: Formulation, Optimization, and In Vitro Characterization. Drug Dev and Ind Pharm. 35, 2009, 583-593
- [53]. Wang JJ, Zeng ZW, Xiao RZ, Xie T, Zhou GL, Zhan XR, Wang SL. Recent advances of chitosan nanoparticles as drug carriers. Int J Nanomed 6, 2011, 765-774.
- [54]. Zhang R, He R, Qian J, Guo J, Xue K, Yuan YF. Treatment of experimental autoimmune uveoretinitis with intravitreal injection of tacrolimus [FK506] encapsulated in liposomes. Invest Ophthalmol Vis Sci. 51, 2010, 3575-3582.
- [55]. Gupta C, Daechsel AK, Chauhan A. Source Interaction of ionic surfactants with cornea-mimicking anionic liposomes. Langmuir. 27, 2011, 10840-10846.
- [56]. Vandamme TF, Brobeck L. Poly [amidoamine] dendrimers as ophthalmic vehicles for ocular delivery of pilocarpine nitrate and tropicamide. J Control Release. 102, 2005, 23-38.
- [57]. Shaunak S, Thomas S, Gianasi E, Godwin A, Jones E, Teo I, Mireskandari K, Luthert P, Duncan R, Patterson S, Khaw P, Brocchini S. Polyvalent dendrimer glucosamine conjugates prevent scar tissue formation. Nat Biotechnol 22, 2004, 977–984.
- [58]. Marano RJ, Toth I, Wimmer N, Brankov M, and Rakoczy PE. Dendrimer delivery of an anti-VEGF oligonucleotide into the eye: a long-term study into inhibition of laser-induced CNV, distribution, uptake and toxicity. Gene therapy. 12, 2005, 1544–1550.
- [59]. Jimenez N, Galan J, Vallet A, Egea MA, Garcia ML. Methyl trypsin loaded poly[D,L-lactide-coglycolide] nanoparticles for contact lens care. J Pharm Sci. 99, 2010, 1414-1426.
- [60]. Vega E, Gamisans F, García ML, Chauvet A, Lacoulonche F, Egea MA. PLGA nanospheres for the ocular delivery of flurbiprofen: drug release and interactions. J Pharm Sci. 97, 2008, 5306-5317.
- [61]. Zhang L, Li Y, Zhang C, Wang Y, Song C. Pharmacokinetics and tolerance study of intravitreal injection of dexamethasone-loaded nanoparticles in rabbits. Int J Nanomedicine. 4, 2009, 175-183.
- [62]. Mandal B, Halder KK, Dey SK, Bhoumik M, Debnath MC, Ghosh LK. Development and physical characterization of chloramphenicol loaded biodegradable nanoparticles for prolonged release. Pharmazie. 64, 2009, 445-449.
- [63]. Sakai T, Kohno H, Ishihara T, Higaki M, Saito S, Matsushima M, Mizushima Y, Kitahara K. Treatment of experimental autoimmune uveoretinitis with poly[lactic acid] nanoparticles encapsulating betamethasone phosphate. Exp Eye Res. 82, 2006, 657-663.
- [64]. Salgueiro A, Egea MA, Espina M, Valls O, García ML. Stability and ocular tolerance of cyclophosphamide-loaded nanospheres. J Microencapsul. 21, 2004, 213-223.
- [65]. Harmia T, Kreuter J, Speiser P, Boye T, Gurny R, Kubi A. Enhancement of the myotic response of rabbits with pilocarpine-loaded polybutylcyanoacrylate nanoparticles. Int J Pharm. 33, 1986, 187-193.
- [66]. Marchal-Heussler L, Sirbat D, Hoffman M, Maincent P. Poly[epsilon-caprolactone] nanocapsules in carteolol ophthalmic delivery. Pharm Res. 10, 1993, 386-390.
- [67]. Das S, Suresh PK, Desmukh R. Design of Eudragit RL 100 nanoparticles by nanoprecipitation method for ocular drug delivery. Nanomedicine. 6, 2010, 318-323.
- [68]. Adibkia K, Omidi Y, Siahi MR, Javadzadeh AR, Barzegar-Jalali M, Barar J, Maleki N, Mohammadi G, Nokhodchi A. Inhibition of endotoxin-induced uveitis by methylprednisolone acetate nanosuspension in rabbits. J Ocul Pharmacol Ther. 23, 2007, 421–432.

- [69]. Enriquez de Salamanca A, Diebold Y, Calonge M, Garcia-Vazquez C, Callejo S, Vila A, Alonso MJ. Chitosan nanoparticles as a potential drug delivery system for the ocular surface: toxicity, uptake mechanism and in vivo tolerance. Invest Ophthalmol. Vis Sci. 47, 2006, 1416–1425.
- [70]. Badawi AA, El-Laithy HM, El Qidra RK, El Mofty H, El Dally M. Chitosan based nanocarriers for indomethacin ocular delivery. Arch Pharm Res. 31, 2008, 1040–1049.
- [71]. De Campos AM, Sanchez A, Alonso MJ. Alonso. Chitosan nanoparticles: a new vehicle for the improvement of the delivery of drugs to the ocular surface. Application to cyclosporine A Int J Pharm. 224, 2001, 159-168.
- [72]. Merodio M, Irache JM, Valamanesh F, Mirshahi M. Ocular disposition and tolerance of ganciclovir-loaded albumin nanoparticles after intravitreal injection in rats. Biomaterials. 2397, 2002, 1587–1594.
- [73]. Das, S Banerjee R, Bellare J. Aspirin Loaded Albumin Nanoparticles by Coacervation: Implications in Drug Delivery. Trends Biomater. Artif. Organs. 18, 2005, 203-212.
- [74]. De Kozak Y, Andrieux K, Villarroya H, Klein C, Thillaye-Goldenberg B, Naud MC, Garcia E, Couvreur P. Intraocular injection of tamoxifen-loaded nanoparticles: a new treatment of experimental autoimmune uveoretinitis. Eur J Immunol. 34, 2004, 3702-3712.
- [75]. Dillen K, Vandervoort J, Van den Mooter G, Ludwig A. Evaluation of ciprofloxacin-loaded Eudragit RS100 or RL100/PLGA nanoparticles. Int J Pharm. 314, 2006, 72-82.
- [76]. Ibrahim HK, El-Leithy IS, Makky AA. Mucoadhesive nanoparticles as carrier systems for prolonged ocular delivery of gatifloxacin/prednisolone bitherapy. Mol Pharm. 7, 2010, 576-585.
- [77]. Parveen S, Mitra M, Krishnakumar S, Sahoo SK. Enhanced antiproliferative activity of carboplatin-loaded chitosan-alginate nanoparticles in a retinoblastoma cell line. Acta Biomater. 6, 2010, 3120-3131.
- [78]. Wadhwa S, Paliwal R, Paliwal SR, Vyas SP. Hyaluronic acid modified chitosan nanoparticles for effective management of glaucoma: development, characterization, and evaluation. J Drug Target. 18, 2010, 292-302.
- [79]. Giannavola C, Bucolo C, Maltese A, Paolino D, Vandelli MA, Puglisi G, Lee VH, Fresta M. Influence of preparation conditions on acyclovir-loaded poly-d,l-lactic acid nanospheres and effect of PEG coating on ocular drug bioavailability. Pharm Res. 20, 2003, 584-590.
- [80]. Tian B, Luo Q, Song S, Liu D, Pan H, Zhang W, He L, Ma S, Yang X, Pan W. Novel surface-modified nanostructured lipid carriers with partially deacetylated.
- [81]. Nagarwal RC, Singh PN, Kant S, Maiti P, Pandit JK. Chitosan coated PLA nanoparticles for ophthalmic delivery: characterization, in-vitro and in-vivo study in rabbit eye. J Biomed Nanotechnol. 6, 2010, 648-657.
- [82]. Luo Q, Zhao J, Zhang X, Pan W. Nanostructured lipid carrier [NLC] coated with Chitosan Oligosaccharides and its potential use in ocular drug delivery system. Int J Pharm. 403, 2011, 185-191.
- [83]. De Campos AM, Sanchez A, Gref R, Calvo P, Alonso MJ. The effect of a PEG versus a chitosan coating on the interaction of drug colloidal carriers with the ocular mucosa, Eur J Pharm Sci. 20, 2003, 73–81.
- [84]. Kalam MA, Sultana Y, Ali A, Aqil M, Mishra AK, Chuttani K. Preparation, characterization, and evaluation of gatifloxacin loaded solid lipid nanoparticles as colloidal ocular drug delivery system. J Drug Target. 18, 2010, 191-204.
- [85]. Li R, Jiang S, Liu D, Bi X, Wang F, Zhang Q, Xu Q. A potential new therapeutic system for glaucoma: solid lipid nanoparticles containing methazolamide J Microencapsul. 28, 2011, 134-141.
- [86]. Liu Z, Zhang X, Wu H, Li J, Shu L, Liu R, Li L, Li N. Preparation and evaluation of solid lipid nanoparticles of baicalin for ocular drug delivery system in vitro and in vivo.. Drug Dev Ind Pharm. 37, 2011, 475-481.
- [87]. Attama AA, Reichl S, Müller-Goymann CC. Sustained release and permeation of timolol from surface-modified solid lipid nanoparticles through bioengineered human cornea. Curr Eye Res. 34, 2009, 698-705.
- [88]. Basaran E, Demirel M, Sirmagül B, Yazan Y. Cyclosporine-A incorporated cationic solid lipid nanoparticles for ocular delivery. J Microencapsul. 27, 2010, 37-47.
- [89]. Cavalli R, Gasco R, Chetoni P, Burgalassi S, Saettone MF. Solid lipid nanoparticles [SLN] as ocular delivery system for tobramycin. Int J Pharm. 238, 2002, 241–245.
- [90]. Hao J, Fang X, Zhou Y, Wang J, Guo F, Li F, Peng X. Development and optimization of solid lipid nanoparticle formulation for ophthalmic delivery of chloramphenical using a Box-Behnken design. Int J Nanomedicine. 6, 2011, 683-692.

- [91]. Attama AA, Reichl S, Müller-Goymann CC. Diclofenac sodium delivery to the eye: in vitro evaluation of novel solid lipid nanoparticle formulation using human cornea construct. Int J Pharm. 355, 2008, 307-313.
- [92]. Ludwig A. The use of mucoadhesive polymers in ocular drug delivery. Adv drug Deliv Rev. 57, 2005, 1595-1639.
- [93]. Calvo P, Vila-JL, Alonso MJ. Evaluation of cationic polymer-coated nanocapsules as ocular drug carriers. Int J Pharm 153, 1997, 41-50.
- [94]. Schipper NG, Olsson S, Hoogstraate JA, deBoer AG, Vårum KM, Artursson P Chitosans as absorption enhancers for poorly absorbable drugs 2: mechanism of absorption enhancement. Pharm Res 14, 1997, 923-929.
- [95]. Du Toit LC, Pillay V, Choonara YE, Govender T, Carmichael T. Ocular drug delivery a look towards nanobioadhesives. Expert Opin Drug Deliv. 8, 2011, 71-94.
- [96]. Hirano S, Seino H, Akiyama I, Nonaka I. Chitosan: a biocompatible material for oral and intravenous administration. In: Gebelein CG, Dunn RL, editors, Progress in biomedical polymers. Plenum Press, New York; 1990, 283-289.
- [97]. Takeuchi H, Yamamoto H, Niwa T, Hino T, Kawashima Y. Enteral absorption of insulin in rats from mucoadhesive chitosan coated liposomes. Pharm Res 13, 1996, 896-901.
- [98]. Felt O, Furrer P, Mayer JM, Plazonnet B, Buri P, Gurny R. Topical use of chitosan in ophthalmology: tolerance assessment and evaluation of pre-corneal retention. Int J Pharm 180, 1999, 185-193.
- [99]. Schoenwald RD, Ward R. Relationship between steroid permeability across excised rabbit cornea and octanol-water partition coefficients. J Pharm Sci 67, 1978, 786-788.
- [100]. Ludwig A, van Haeringen NJ, Bodelier VM, Van Ooteghem M. Relationship between precorneal retention of viscous eye drops and tear fluid composition. Int. Ophthalmol. 16, 1992, 23-26.
- [101]. Van Ooteghem M. Preparations ophtalmiques. In: Galenica, editor, Technique and documentation. Lavoisier, Paris; 1995.
- [102]. Greaves JL, Olejnik O, Wilson CG. Polymers and the precorneal tear film. S.T.P. Pharma Sciences. 2, 1992, 13-33.
- [103]. Ponchel G, Touchard F, Duchfine D, Peppas NA. Bioadhesive analysis of controlled-release systems. I. Fracture and interpenetration analysis in poly [acrylic acid]-containing systems. J Control Release 5, 1987, 129-141.
- [104]. Park H, Robinson JR. Mechanisms of mucoadhesion of poly[acrylic acid] hydrogels. Pharm Res. 4, 1987, 457-464.
- [105]. Slovin EM, Robinson JR. Bioadhesives in ocular drug delivery. In: Edman P, editor, Biopharmaceutics of ocular drug delivery. CRC Press, Boca Raton, Florida; 1993, 145-57.
- [106]. Duchfine D, Touchard F, Peppas NA. Pharmaceutical and medical aspects of bioadhesive systems of drug administration. Drug Dev Ind Pharm 14, 1988, 283-318.
- [107]. Barbault-Foucher S, Gref R, Russo P, Guechot J, Bochot A.. Design of poly–caprolactone nanospheres coated with bioadhesive hyaluronic acid for ocular delivery. J Control Release 83, 2002, 365-375.
- [108]. Lemarchand C, Gref R, Couvreur P. Polysaccharide-decorated nanoparticles. Eur J Pharm Biopharm 58, 2004, 327-341.
- [109]. Sintzel MB, Bernatchez SF, Tabatabay C, Gurny R. Biomaterials in ophthalmic drug delivery. Eur J Pharm Biopharm 42, 1996, 358-374.
- [110]. Friess W. Collagen-material for drug delivery. Eur J Pharm Biopharm 45, 1998, 113-136.
- [111]. Vandervoort J, Ludwig A. Preparation and evaluation of drug-loaded gelatin nanoparticles for topical ophthalmic use. Eur J Pharm Biopharm. 57, 2004, 251-261.
- [112]. Guo LSS, Sarris AM, Levy MD. A safe bioadhesive liposomal formulation for ophthalmic applications. Invest Ophthalmol Vis Sci 29, 1989, 439.
- [113]. Calvo P, Alonso MJ, Vila-Jato JL, Robinson JR. Improved ocular bioavailability of indomethacin by novel ocular drug carriers. J Pharm Pharmacol. 48, 1996, 1147–1152.
- [114]. De la Fuente M, Seijo B, Alonso MJ. Bioadhesive hyaluronan–chitosan nanoparticles can transport genes across the ocular mucosa and transfect ocular tissue, Gene Ther. 15, 2008, 668–676.

- [115]. Yuan XB, Yuan YB, Jiang W, Liu J, Tian EJ, Shun HM, Huang DH, Yuan XY, Li H, Sheng J. Preparation of rapamycin-loaded chitosan/PLA nanoparticles for immunosuppression in corneal transplantation, Int J Pharm 349, 2008, 241–248.
- [116]. Qu X, Khutoryanskiy VV, Stewart A, Rahman S, Papahadjopoulos-Sternberg B, Dufes C, McCarthy D, Wilson CG, Lyons R, Carter KC, Schätzlein A, Uchegbu IF. al. Carbohydrate-based micelle clusters which enhance hydrophobic drug bioavailability by up to 1 order of magnitude. Biomacromolecules 7, 2006, 3452–3459.
- [117]. Aggarwal D, Kaur IP. Improved pharmacodynamics of timolol maleate from a mucoadhesive niosomal ophthalmic drug delivery system. Int J Pharm. 290, 2005, 155-159.
- [118]. Kaur IP, Singh M, Kanwar M. Formulation and evaluation of ophthalmic preparation of acetazolamide. Int J Pharm. 199, 2000, 119–127.